



Real time RT-PCR normalisation: methods and considerations.

Dr. Jim Huggett

Centre for Infectious Diseases
University College London
Windeyer Building
46 Cleveland Street
London, W1T 4JF

j.huggett@ucl.ac.uk

Research projects at the Center for Infectious Diseases & International Health in the Developing World

- **Clinical trials (Drugs, Vaccine, Diagnostic tests)**
- **Epidemiological studies**
- **Basic science research**



- **Basic science research**

 - **Immunology, Biochemistry, Microbiology, Genetics, Molecular biology**

- **Training/ Laboratory setup**

- **Troubleshooting, Support**

- **Assay development**



Normalisation of real time RT-PCR

- **Real time PCR can be very accurate over a large dynamic range**
- **Measuring RNA by RT-PCR accuracy can be effected by error:**
 - **Inherent variability of RNA**
 - **Multi-step process required for analysis**
- **Normalisation required to control for this error**

Normalisation of real time RT-PCR

Sample

Extract RNA ↓ ← *Ensure similar sample size*

RNA

Generate cDNA ↓ ← *Ensure similar RNA concentration*

cDNA

Measure cDNA by
Real time PCR ↓ ← *Measure internal reference*

Result

Sample

Extract RNA ↓ ← *Ensure similar sample size*

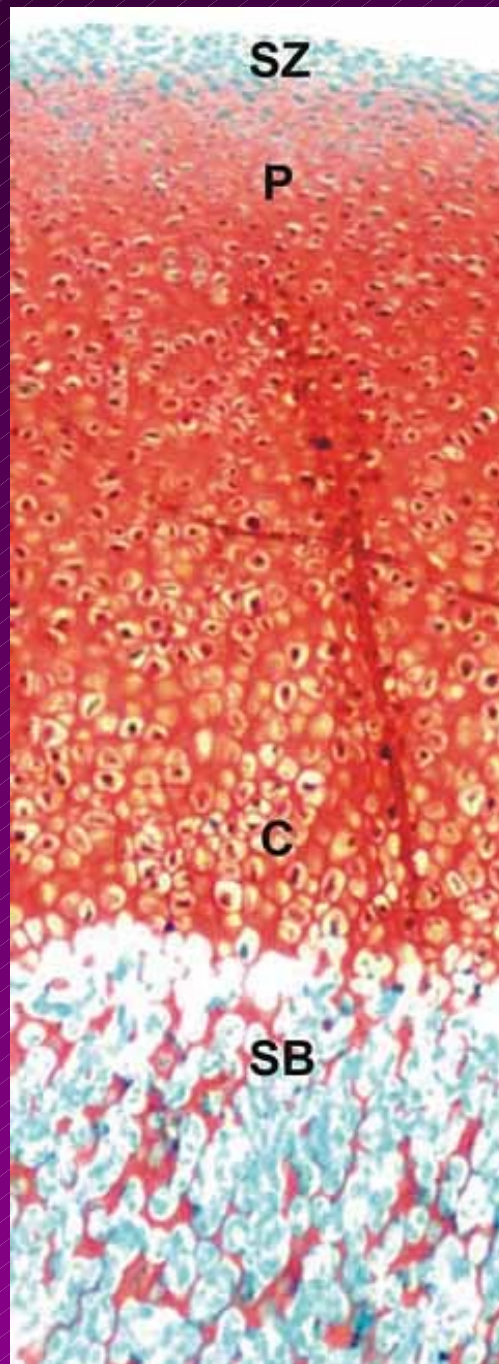
RNA

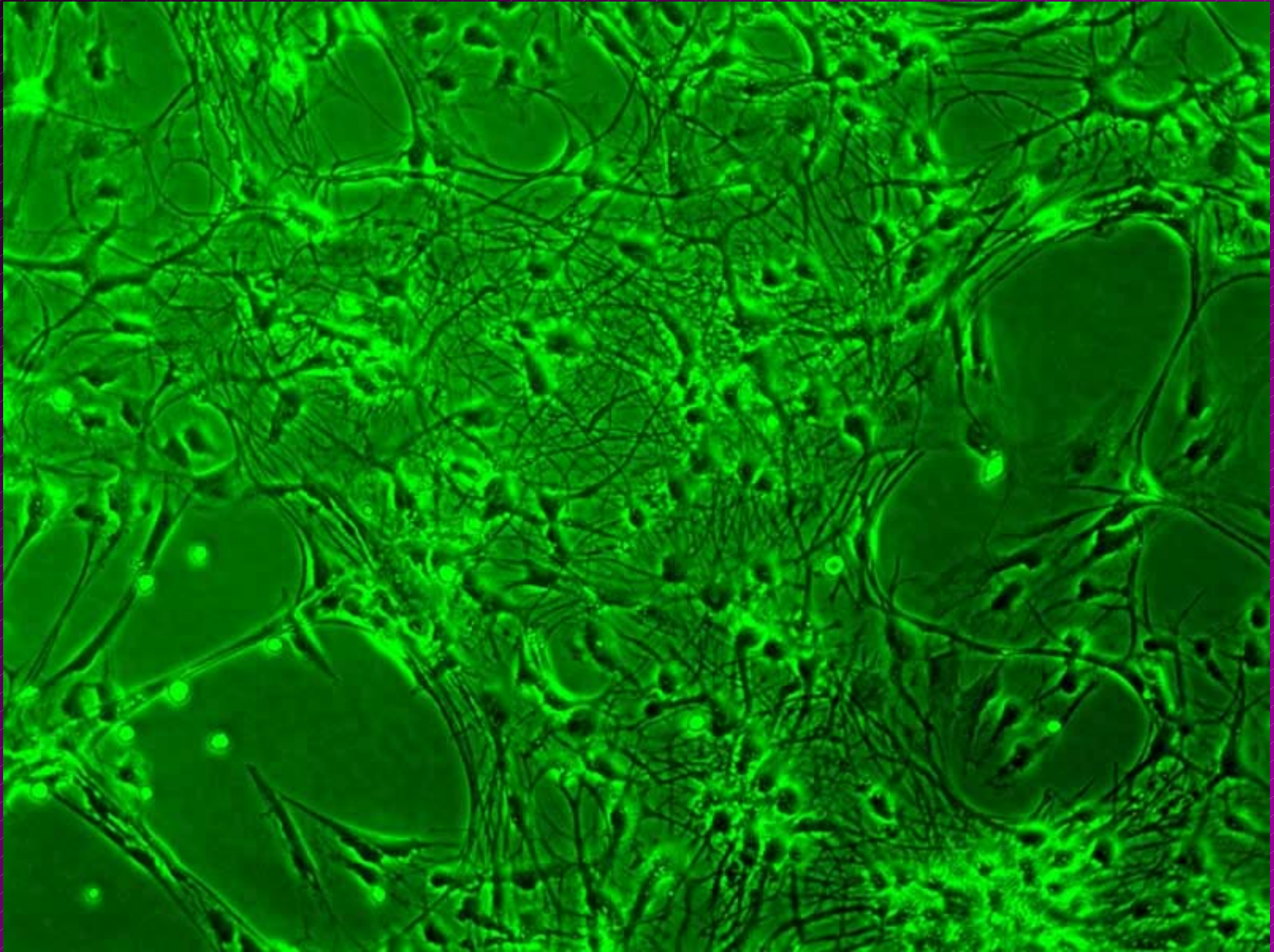
Generate cDNA ↓

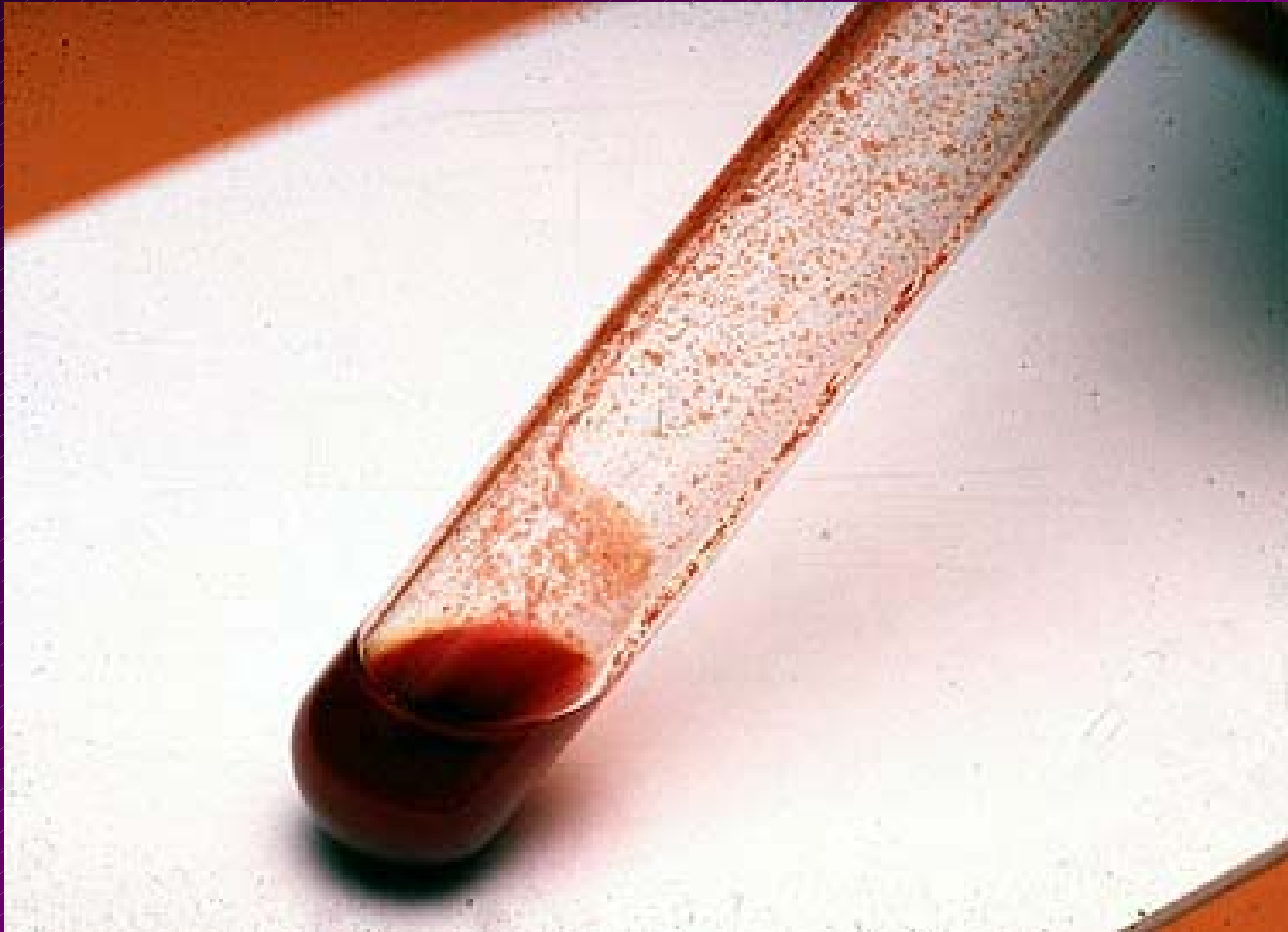
cDNA

Measure cDNA by
Real time PCR ↓

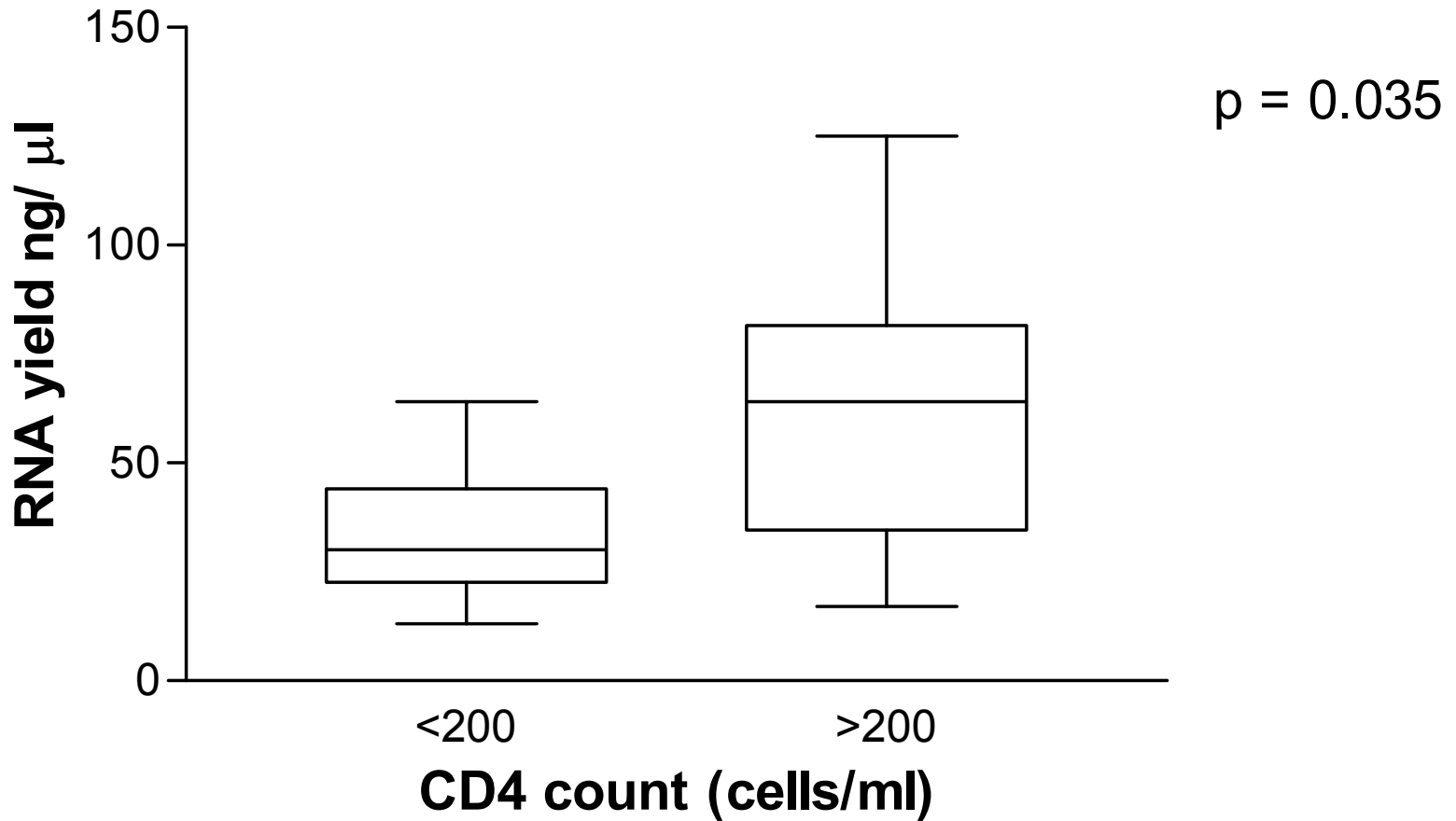
Result







Total RNA extracted



Sample

Extract RNA ↓ ← *Ensure similar sample size*

RNA

Generate cDNA ↓ ← *Ensure similar RNA amount*

cDNA

Measure cDNA by
Real time PCR ↓

Result

Normalising to [RNA]

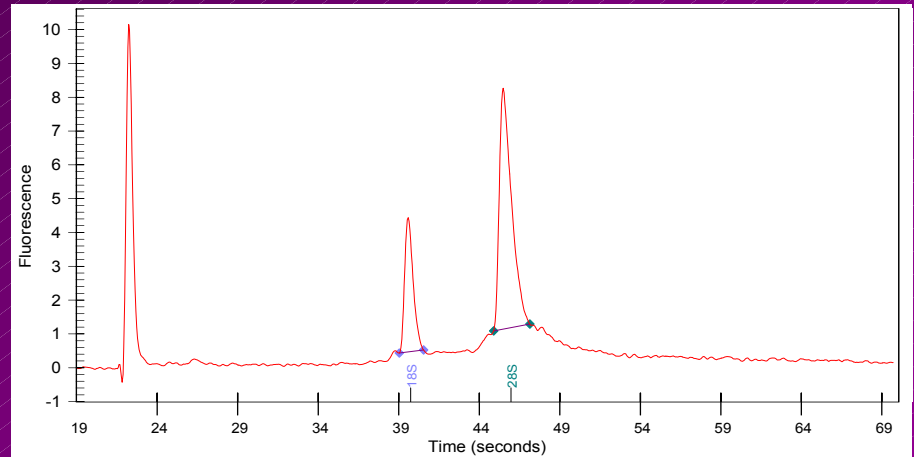
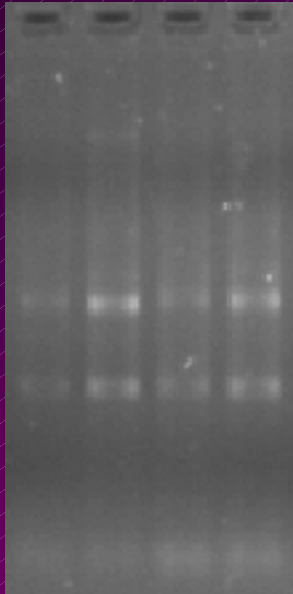
- **UV (OD_{260})**
- **Gel analysis (Agarose, PAGE, Bioanalyser)**
- **Ribo-green**

Normalising to [RNA]

- Quantity
- Quality

Normalising to [RNA]

- Quality



Normalising to [RNA]

- **Does not control for error introduced in subsequent steps**
- **Assumes no change in rRNA:mRNA ratio**

Sample

Extract RNA   *Ensure similar sample size*

RNA

Generate cDNA   *Ensure similar RNA concentration*

cDNA

Measure cDNA by
Real time PCR   *Measure internal reference*

Result

Internal reference

- **Reference Gene (House Keeping Gene)**
- **Artificial Spike**
- **Genomic DNA**

Reference genes

Genes that are assumed to be expressed at a constant level between experimental groups

Reference genes

- **Originate from RNA qualitative/semi-quantitative methods**
- **Examples:**
 - **glyceraldehyde-3-phosphate dehydrogenase (GAPDH)**
 - **β -actin**
 - **hypoxanthine-guanine phosphoribosyl transferase (HPRT)**
 - **18S rRNA**
- **Ubiquitous RNAs are ideal positive controls**
- **BUT are they constitutively expressed?**

Reference genes

- **real time RT-PCR made RNA measurement quantitative over a huge dynamic range**
- **Surely when this happened the scientific world would have checked that reference were actually constitutively expressed.**

- Tanaka *et al.* 1975 18S rRNA was reported to increase in expression with cytomegalovirus infection.
- In 1984 Piechaczyk *et al.* reported that while GAPDH transcription occurred at a similar rate in different rat tissues they contained very different amounts of mRNA.
- In 1985 Stout *et al.* HPRT is constitutively expressed at low levels in most human tissues but was elevated in certain parts of the central nervous system
- In 1987 Blomberg *et al.* β -actin mRNA differentially expressed in different leukaemia patient tumour samples.

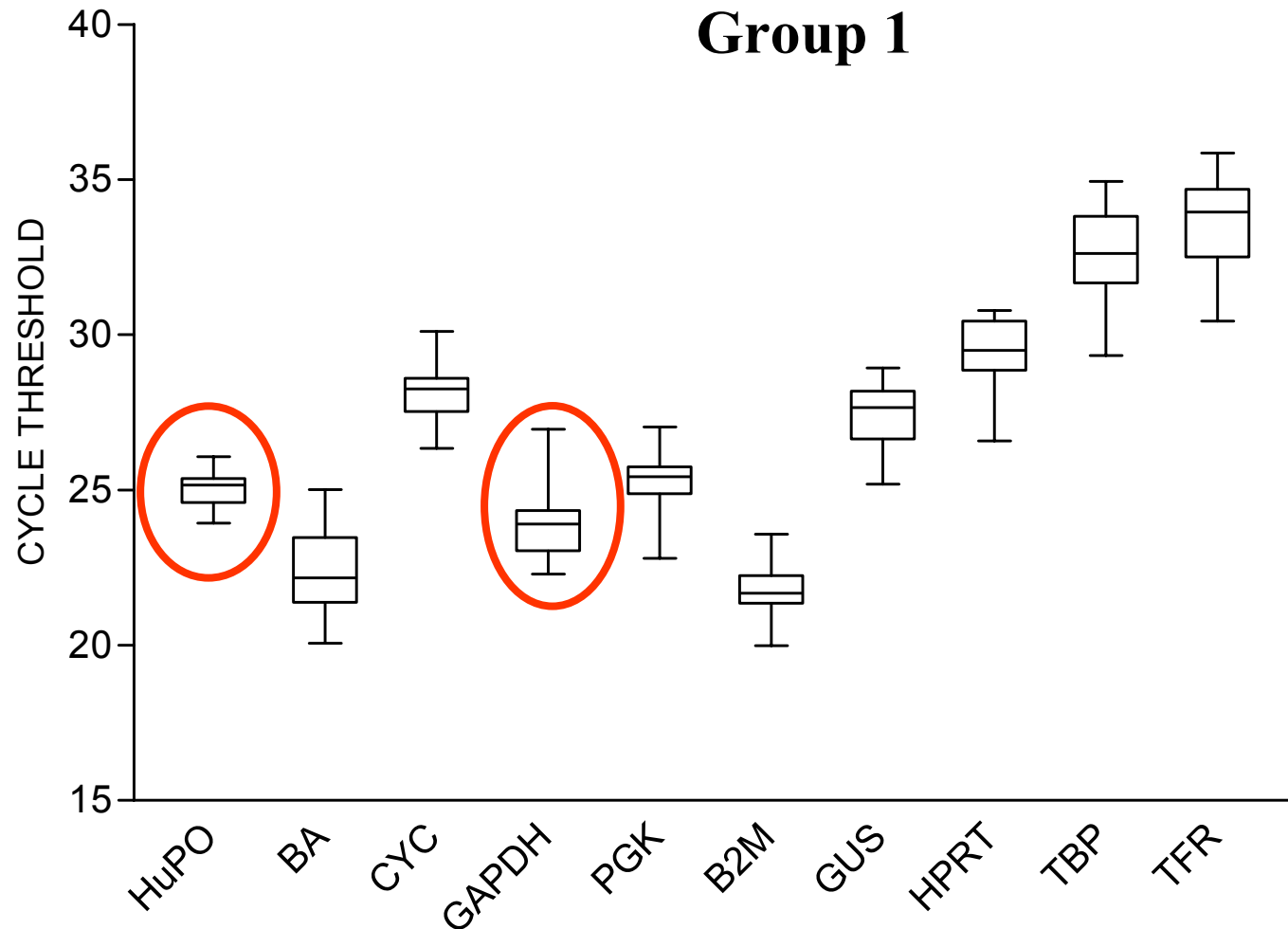
- The notion that these RNAs require validation is also not new.
- 1989 Barbu *et al.* reported that probes to β -actin were inappropriate as a reference when comparing different mouse tissue by northern blot

**There is a lot of published data
that has used invalidated
reference genes for
normalisation**

- Study investigating the immunology of Tuberculosis
- We used the Taqman Human Endogenous Control Plate to investigate 10 potential house keeping genes

Dheda K, Huggett JF, Bustin SA, Johnson MA, Rook G, Zumla A.
Biotechniques. 2004 Jul;37(1):112-4, 116, 118-9.

CT Variation of house keeping genes from human blood



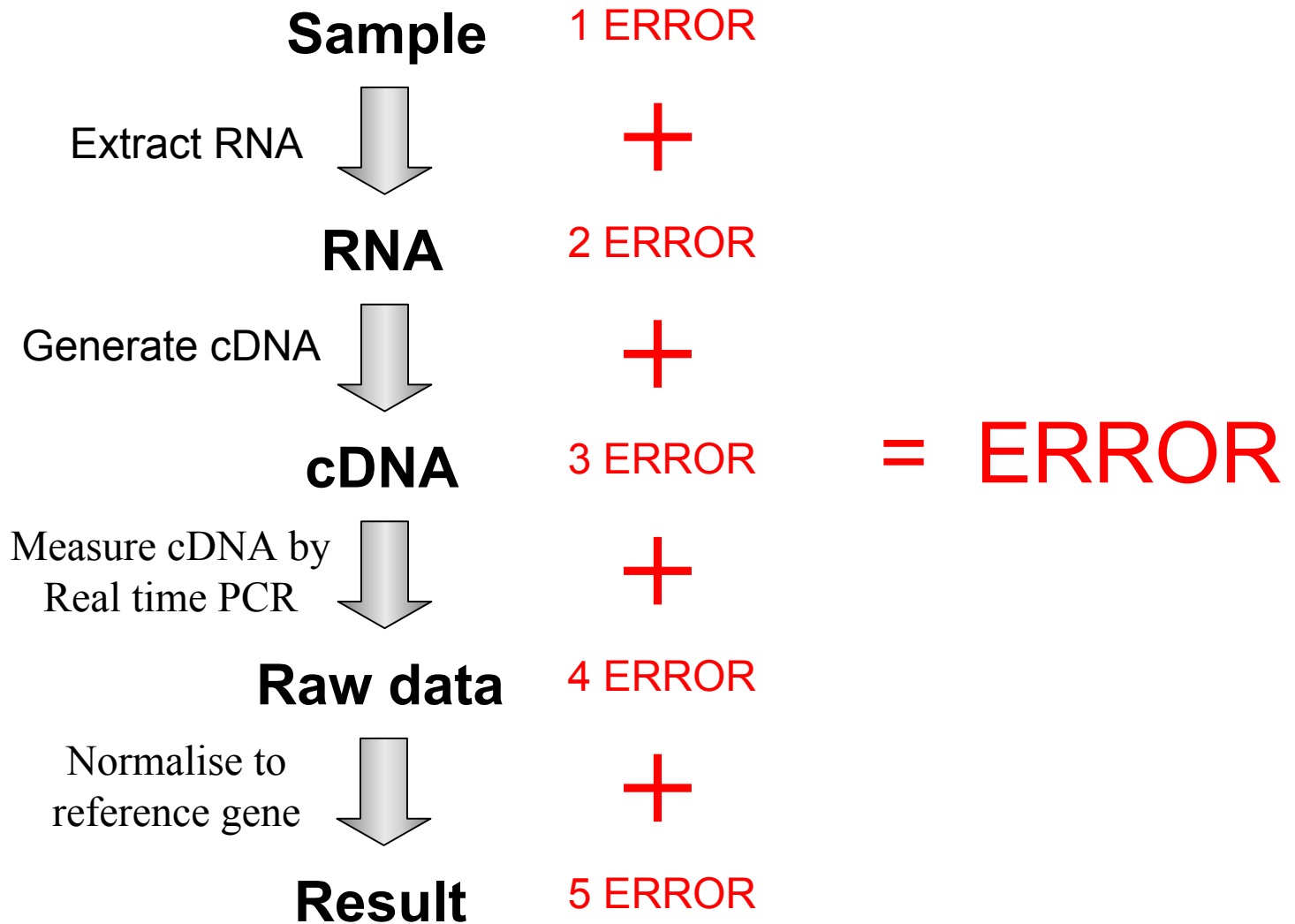
Median, 25th & 75th percentile and range

2 things to measure with reference gene validation

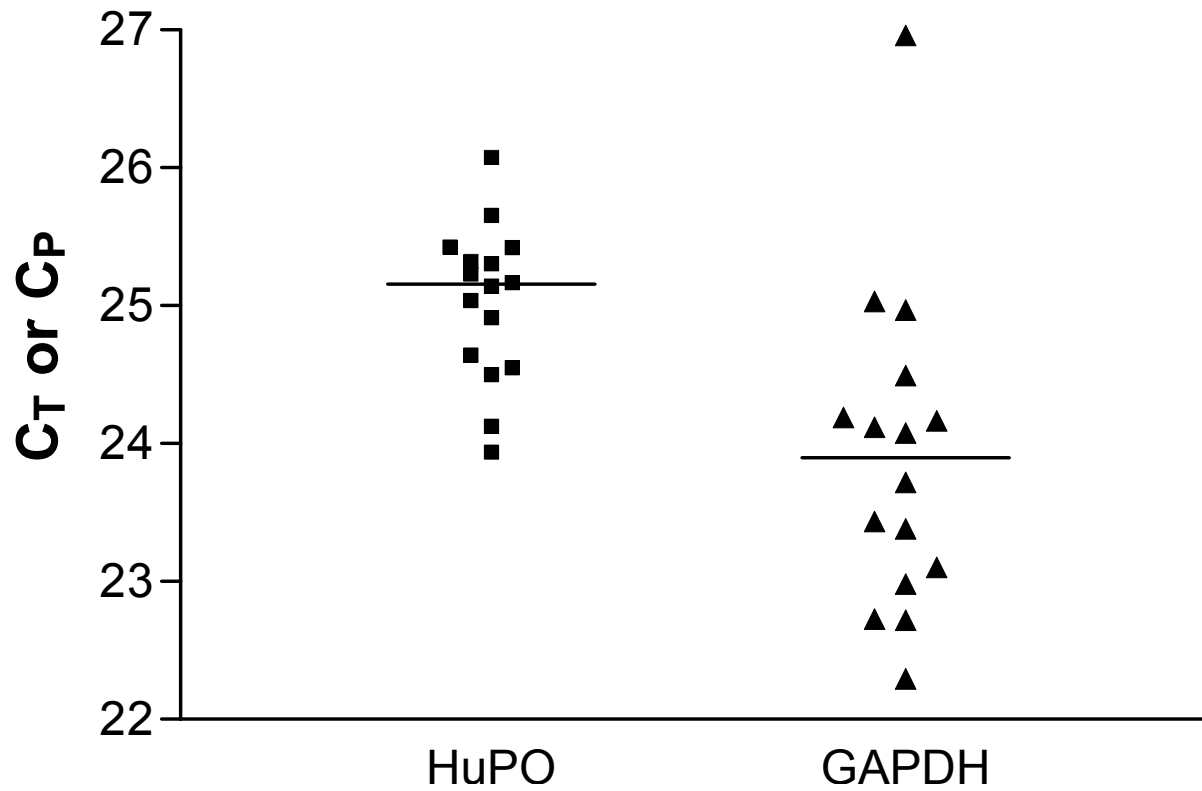
1. General error

2. Directional error (directional shift)

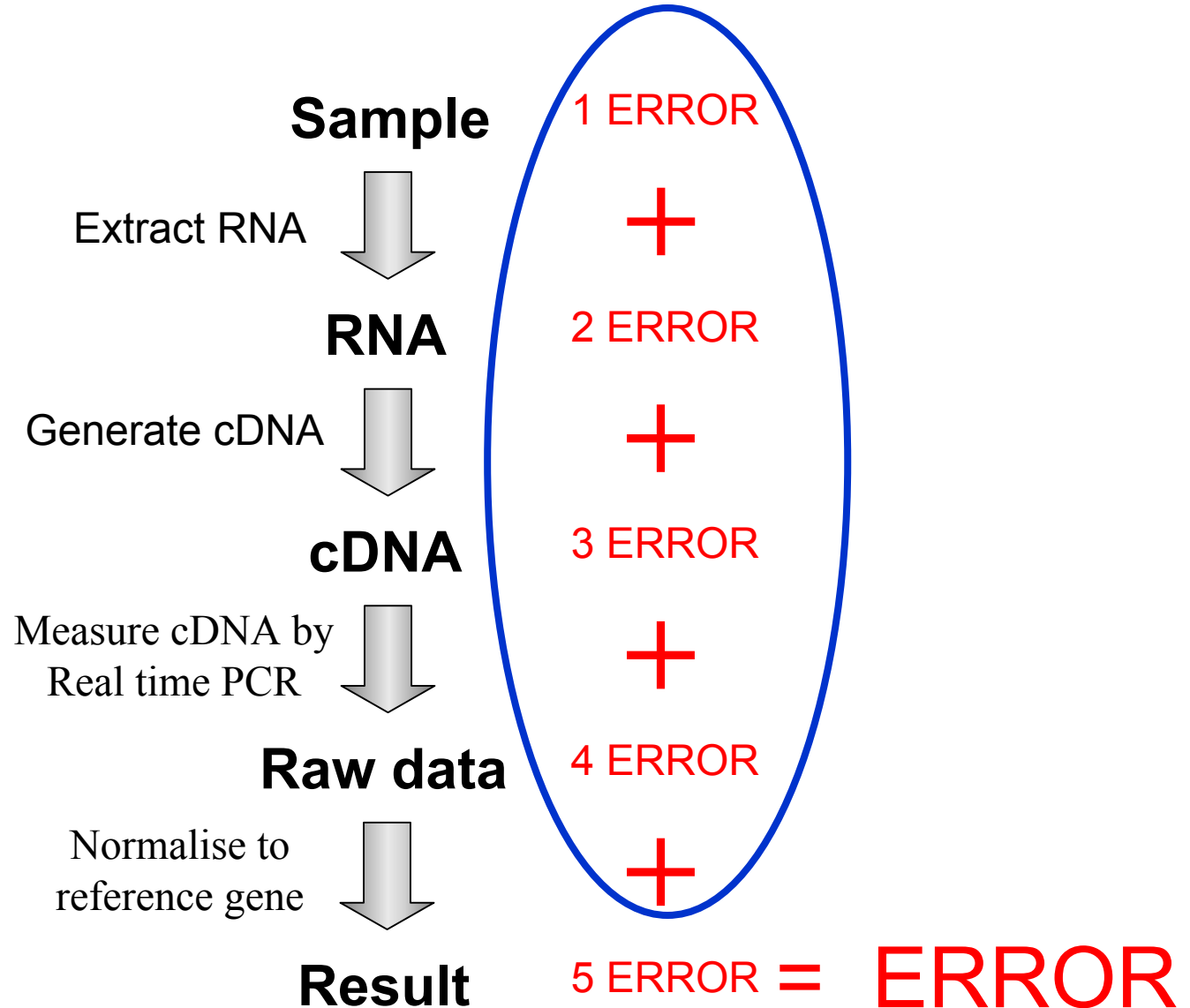
1. General error



1. General error



1. General error



1. General error

- The error of the chosen reference gene defines the resolution of the assay
- This resolution is dependent on the desired measurement

i.e. 1 log variation in reference gene is OK for a 2 log measurement

2. Directional Shift

- Assumption reference gene constitutive expression
- Variation reference gene expression is always random
- What if the experiment effects chosen reference?

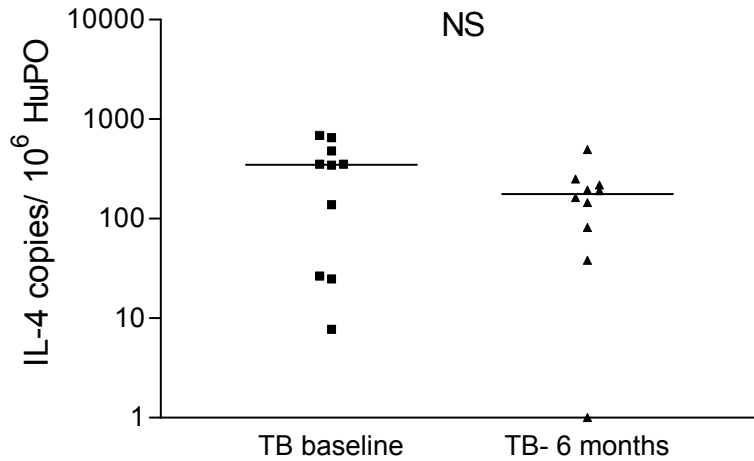
2. Directional shift effect: example 1

- Comparison of Interleukin 4 (IL4) mRNA expression between Tuberculosis patients and healthy individuals.
- Normalise raw data to HuPO or GAPDH

2. Directional shift effect: example 2

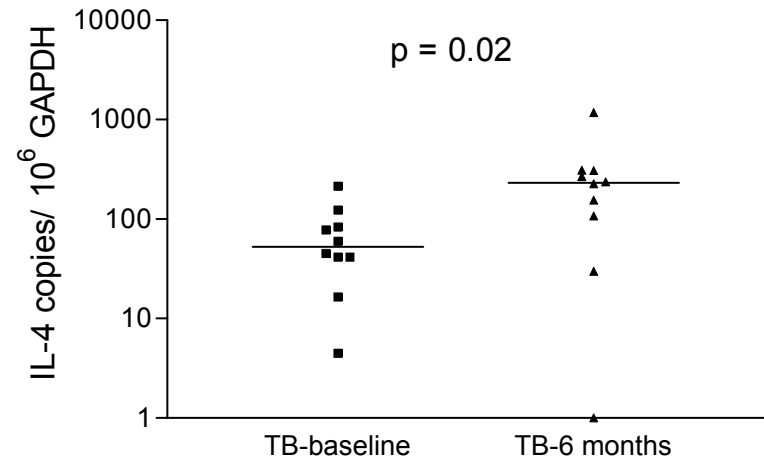
- Effect of treatment on IL4 mRNA expression
- Normalise raw data to HuPO or GAPDH

2. Directional shift



Normalised to 10⁶ copies HuPO

Normalised to 10⁶ copies GAPDH



Conclusion

- **There are a number of methods for normalising real time RT-PCR experiments which are not mutually exclusive**
- **Using a reference gene is a simple and effective method**
- **However it must be validated for the specific experiment using the actual samples being measured**
- **Failure to do so may generate incorrect results**

Acknowledgements

•Center for Infectious Diseases and International Health,
University College London

Prof. Zumla

Dr. Kim

Prof. Rook

Dr. Dheda

Miss Chang

•Queen Mary's School of Medicine, University of London
Prof. Bustin

•Funding.
DFID, EU, EDCTP, BLF